

NUCLEAR MAGNETIC RESONANCE SPECTROSCOPY.
THE STRUCTURE OF BUTENYLMAGNESIUM
BROMIDE¹

Sir:

Crotyl bromide and α -methylallyl bromide each react with magnesium in ether to yield the same butenyl Grignard reagent, which conceivably could be crotylmagnesium bromide (I), α -methylallyl-

reactions, Young and Roberts^{2a} inferred that the correct structure was crotylmagnesium bromide (I). This view was sharply criticized by Kharasch and Reinmuth,⁴ who held that the problem could be solved only by physical means. The ultimate justice of the criticism is not to be denied; nonetheless, infrared evidence⁵ and the nuclear magnetic resonance (n.m.r.) spectrum, as will be shown below,

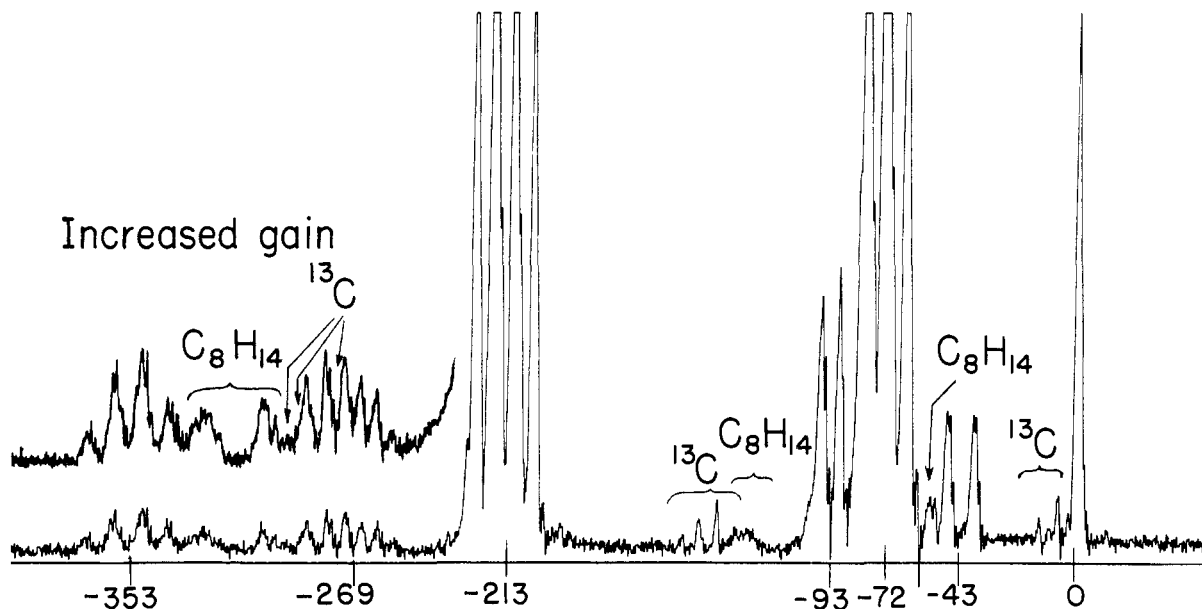
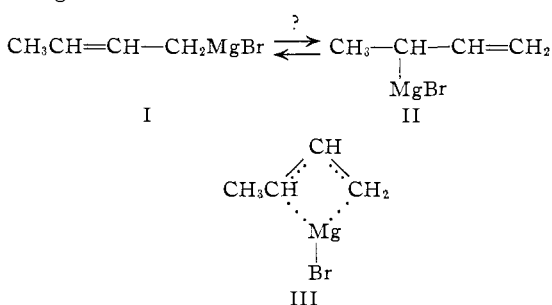


Fig. 1.—Nuclear magnetic resonance spectrum of butenylmagnesium bromide in diethyl ether at 25° and 60 Mc with tetramethylsilane (0 cps.) as internal standard. The intense triplet and quartet resonances centered on -72 and -213 cps. are the CH₃ and CH₂ of diethyl ether. The lines designated as ¹³C are the proton satellite resonances produced by the ¹³C in the solvent. The resonances arising from the octadienes formed by coupling in the preparation of the Grignard reagent are marked C₈H₁₄. The assignments of these were verified by addition of authentic material to the sample. The resonances of the Grignard protons were assigned as follows: α -CH₂ doublet centered on -43 cps.; δ -CH₃, doublet centered on -93 cps.; γ -CH, quintet centered on -269 cps.; and β -CH, quartet centered on -353 cps. The spin-spin splitting pattern is discussed in detail elsewhere.⁸

magnesium bromide (II), a more or less mobile equilibrium mixture of I and II, or an intermediate, bridged structure such as III.^{2,3}



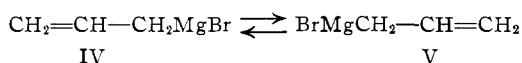
On the basis of the extraordinary behavior of the butenyl Grignard reagent in a variety of chemical

strongly support the inference that the butenyl Grignard reagent, at least largely and probably exclusively, is the crotyl isomer.

The n.m.r. spectrum of the butenyl Grignard reagent is independent of whether or not one starts from crotyl or α -methylallyl bromide and is shown in Fig. 1. The spectrum is that expected for I since there are two vinyl proton resonances and an upfield two-proton resonance that is split into a doublet as expected for a -CH₂Mg- group connected to a carbon carrying a single proton.⁶ The structural problem posed by the Grignard reagent might well be regarded as solved with this finding. However, allylmagnesium bromide has been found to give an n.m.r. spectrum that indicates the allylic isomers IV and V are interconverted faster than 1000 times per second.⁷ A similarly rapid equilibrating mixture of I and II could lead to the spectrum of Fig. 1, provided only that the equilib-

(1) Supported in part by the Office of Naval Research.
(2) (a) W. G. Young and J. D. Roberts, *J. Am. Chem. Soc.*, **68**, 1472 (1946); (b) the chemistry of allylic Grignard reagents is summarized briefly by R. H. de Wolfe and W. G. Young, *Chem. Revs.*, **56**, 753 (1956).
(3) For the purposes of this paper, Grignard reagents will be expressed by the conventional formula RMgX, although there is good reason to believe that this formula is incorrect or, at least, incomplete; cf. R. E. Dessy, G. S. Handler, J. H. Wotiz and C. A. Hollingsworth, *J. Am. Chem. Soc.*, **79**, 3476 (1957); R. E. Dessy and G. S. Handler, *ibid.*, **80**, 5824 (1959).

(4) M. S. Kharasch and O. Reinmuth, "Grignard Reactions of Non-metallic Substances," Prentice-Hall, Inc., New York, N. Y., 1954, Chap. XVII, particularly pages 1154-1157.
(5) M. Gaudemar, *Bull. soc. chim. France*, 1475 (1958).
(6) J. D. Roberts, "Nuclear Magnetic Resonance Spectroscopy," McGraw-Hill Book Company, New York, N. Y., 1959, Chap. III.
(7) J. E. Nordlander and J. D. Roberts, *J. Am. Chem. Soc.*, **81**, 1769 (1959).



rium lies fairly far on the side of I. The extent of the contribution of II to the spectrum determines the chemical shift of the $-\text{CH}_2-$ resonance; if II were the dominant form this resonance would appear in the vinyl region. Since it actually appears in or near the $-\text{CH}_2-\text{Mg}$ region, we can safely infer that I is the dominant form. A comparison⁸ of the effects of vinyl and methyl groups (as R) on the chemical shift of the methylene resonances in $\text{RC}-\text{H}_2\text{X}$, $\text{X} = \text{Br}$, and MgBr suggests that the $-\text{CH}_2-$ resonance of the Grignard reagent in Fig. 1 may actually be some 50 cps. (at 60 Mc) downfield from the position it would have if the Grignard reagent were exclusively crotylmagnesium bromide. This figure corresponds to about 84% of I and 16% of II. A rather strong argument against the validity of the composition so calculated is provided by the fact that the position of the $-\text{CH}_2-$ resonance changes by only 1 cps. between $+30^\circ$ and -20° , corresponding to an increase of but 0.3% in the proportion of the crotyl isomer. It could be argued that through coincidence ΔH is virtually zero for the equilibrium, but it seems more reasonable to conclude that I is more than 99% of the mixture. If so, then even a several-fold temperature effect on the concentration of II would give only a very small change in spectrum.

As with allylmagnesium bromide,⁸ the spectrum of butenylmagnesium bromide shows no evidence of "freezing out"⁹ of the separate allylic isomers at temperatures down to -60° . Interestingly, dibutenylmagnesium shows the same chemical shifts as does the ordinary Grignard reagent. This fact can be taken as an argument against the Grignard formula RMgBr since the bromine ought to have some influence on the chemical shift of $-\text{CH}_2-$ resonance. The n.m.r. evidence gives no support for structure III for the Grignard reagent. Bridged structures are favored for crotyl palladium chloride complex¹⁰ and crotyl cobalt tricarbonyl,¹¹ which have very different n.m.r. spectra from the Grignard reagent.

If the Grignard reagent is accepted to be crotylmagnesium bromide (I) with the possibility of very rapid equilibration with a small proportion of α -methylallylmagnesium bromide (II), then the remaining problem is to account for the high yields of products in which it reacts, even with highly sterically-hindered carbonyl compounds, to insert α -methylallyl groups. Presumably if equilibration were rapid, formation of the less-crowded crotyl addition products would be favored. This dilemma is resolved if we assume that coordination of the carbonyl oxygen with the magnesium of I diminishes the electrophilic character of the magnesium sufficiently to cause the rate of allylic isomerization to be much less than the rate of addition. In these circumstances, only the α -methylallyl group would

be inserted by the cyclic addition mechanism.^{2a} If insertion of the α -methylallyl group were very highly retarded by steric hindrance, as with di-*t*-butyl ketone,¹² then allylic isomerization could lead to the crotyl-addition product.

The n.m.r. spectra of other allylic Grignard reagents will be reported in later papers.

(12) K. W. Wilson, J. D. Roberts and W. G. Young, *THIS JOURNAL*, **72**, 218 (1950).

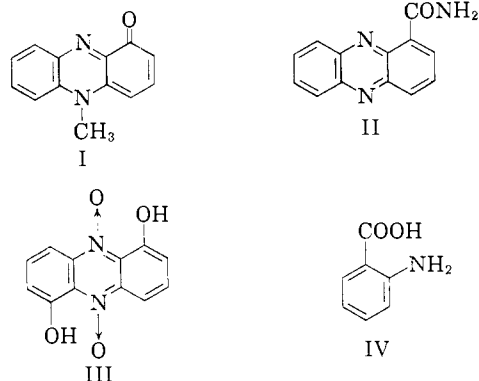
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THE BIOGENESIS OF PHENAZINE PIGMENTS¹

Sir:

Pigments that contain the aromatic phenazine nucleus such as spycocyanine (I),^{1a} chlororaphin (a 3:1 molecular compound of phenazine-1-carboxamide (II) and its 9,10 dihydro derivative),² iodinin (III)³ and phenazine-1-carboxylic acid⁴ present an interesting biogenetic problem for which no definitive proposals have yet been made though a significant amount of work on the biosynthesis of pyrocyanine (I) has been reported.^{5,6,7,8}



We were attracted by the possibility that natural products of this type may arise by an oxidative dimerization of anthranilic acid (IV) with loss of the carboxyl carbon where appropriate. A somewhat tenuously analogous mechanism apparently produces the phenoxazine ring system of the actinomycins.⁹ We wish to report results which we interpret as indicating that one ring, at least, of phenazine-1-carboxamide (II) originates in anthranilic

(1) Supported in part by Grant E-2775 from the U. S. Public Health Service.

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(5) M. V. Burton, J. J. R. Campbell and B. A. Eagles, *Can. J. Res.*, **26C**, 15 (1948).

(6) A. C. Blackwood and A. C. Neish, *Can. J. Microbiol.*, **3**, 165 (1957).

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(9) E. Bullock and A. W. Johnson, *J. Chem. Soc.*, 1602 (1957).

(8) J. E. Nordlander, Ph.D. Thesis, California Institute of Technology, 1960, pp. 41-43.

(9) Ref. 6, Chap. IV.

(10) H. C. Dehn and J. C. W. Chien, *J. Am. Chem. Soc.*, **82**, 4429 (1960).

(11) D. W. Moore, H. B. Jonassen, T. B. Joyner and A. J. Bertrand, *Chem. and Ind.*, 1304 (1960).